

Detection and Enumeration of Pathogenic *Staphylococci* Membrane Filtration Method

Standard XP T90-412 (2006 June)

PREPARATION

Filter 100 ml of water sample on a 0.45 µm sterile membrane

INOCULATION

Apply the membrane on one of the three media below

CHAPMAN

Choose white typical colonies or yellow surrounded with a yellow halo, with the exception of big mucous colonies.

Make a microscopic examination to check the Gram+ cocci appearance.

Mannitol Salt Agar -Chapman
(ref. AEB120511 - 10 x 55 mm plates)

OR

BAIRD PARKER

Choose grey or black typical colonies
Make a microscopic examination to check the Gram+ cocci appearance.

OR

BAIRD PARKER RPF

Make a first reading at (21 ± 3) hours.
Choose grey or black typical colonies with an opacification area on the media under the membrane (presence of coagulase)
Make a microscopic examination to check the Gram+ cocci appearance.

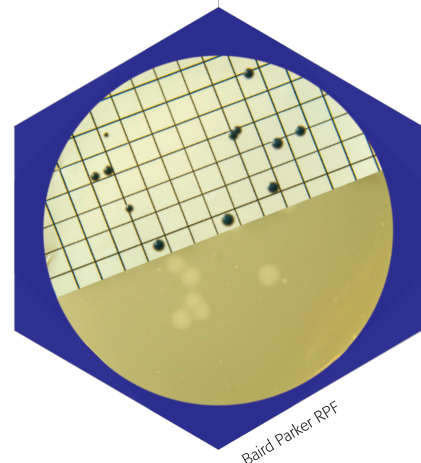
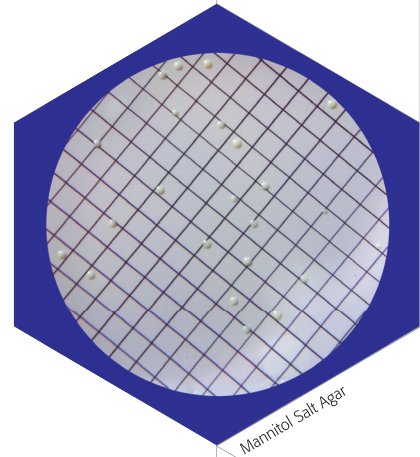
Baird Parker RPF
(ref. AEB120331 - 10 x 55 mm plates)

INCUBATION

Incubate the plate at (36 ± 2)°C during (44 ± 4) hours

- Prick out each colony to be tested in a tube of 10 ml of **Brain Heart Infusion Broth** (ref. AEB110110 – 100 x 10 ml tubes)
Incubate (21 ± 3) hours at (36 ± 2)°C and Perform a coagulase test on each tube - **Rabbit plasma** (ref. 55181 – 8 x 0.5 ml)
- Or a latex agglutination test - **Slidex Staph-kit** (ref.73112 – 50 tests)
- Or a streaking (or a membrane transfer) on **Baird Parker RPF** (ref. AEB120331 - 10 x 55 mm plates) and incubate at (36 ± 2)°C during (21 ± 3) hours

Count as pathogenic *Staphylococci* all colonies appearing as Gram+ cocci and showing a positive coagulase.



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